

Computational Study of Bufadienolides from Indonesia's *Kalanchoe Pinnata* as Na⁺/K⁺-ATPase Inhibitor for Anticancer Agent

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ABSTRACT

Objectives: This study aimed to predict the binding pose of bufadienolides from *Kalanchoe pinnata* on Na⁺/K⁺-ATPase using molecular docking in comparison with a known inhibitor, and to determine the important functional group of bufadienolide that is responsible for good binding. **Material and methods:** Docking was performed on the receptor file (PDB ID: 4RES) using AutoDock 4.2. Before docking, the complex structure of ligand-receptor was minimized using AMBER 14. The physico-chemical properties were predicted using Biovia Draw. **Results:** The sterical hindrance of Glu117 to the 1,3,5-orthoacetate moiety was removed after minimisation. The docking energy, Polar Surface Area (PSA) and A Log P showed good correlation with the experimental data. The 14-OH of all bufadienolides formed a hydrogen bond with Thr797. However, the energy breakdown analysis of docking result demonstrated that **C1** has the highest ligand efficiency with the strongest hydrogen bond and electrostatic energies, followed by **C2** and **C3**. The hydrogen bond from the 10-OH and the orthoacetate ring in **C1** was stronger than that of 10-CH₂OH in **C2** and 10-OH in **C3**. **Conclusion:**

The best activity of **C1**, as compared to **C2** and **C3**, was due to the presence of 1,3,5-orthoacetate moiety, 10-CHO, 11-OH, and 14-OH. In addition, the 11-OH group appeared to decrease the toxicity of the bufadienolide by improving its selectivity to the receptor. This result is expected to be useful in the further development of bufadienolide-based inhibitor for anticancer.

Key words: Bufadienolide, *Kalanchoe Pinnata*, Anticancer, Na⁺/K⁺-ATPase, Molecular Docking.

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INTRODUCTION

Cardiac Steroid (CS) is one of the promising anticancer.¹⁻⁸ The previous study showed that CS exhibited the most potent antitumor activity among 9,000 screened compounds.² In the last decade, there is an increasing number of studies that reported the anticancer activities of CS.^{1,3-6} CS inhibited the proliferation and migration of cells, and also sensitised the multidrug resistant (MDR) strain.⁷ Moreover, Wei and colleagues reported that CS showed significant inhibition tumor growth *in vivo* below their lethal dose.⁸ One of the CS derivatives is bufadienolide. It has an extended six-membered lactone ring at position 17 of the steroidal scaffold. Different with the structure of cholesterol which is all in *trans*-conformation, the ring A/B and C/D in bufadienolides are in *cis*-fused conformation Figure 1. The *cis*-conformation was suggested as the pharmacophore of bufadienolides.⁴ Previously, bufadienolide was used as a treatment for heart disorder disease. However, bufadienolide has also been known to control the growth of cancer cells.^{9,10} One of the proposed molecular mechanisms of its anticancer activity is through the inhibition of Na⁺/K⁺-ATPase (NaK-ATPase).⁹⁻¹⁴ Supratman *et al.* reported the sub-micromolar inhibitory activities of three isolated bufadienolides from Indonesian plant *Kalanchoe Pinnata*, namely **C1-C3** Figure 1, on the tumor-promoted Raji cell line.¹⁵ Computational methods, such as molecular docking, have been used to predict the molecular mechanism of bioactive compounds at the atomic-level.¹⁶⁻¹⁹ Therefore, the aims of this work were to study the structure-activity relationship of NaK-ATPase inhibition of three bufadienolides using molecular docking method.

METHODS

Preparation of Receptor and Ligands

The crystal structure of NAK-ATPase in complex with bufalin was retrieved from Protein Data Bank (rcsb.org/pdb/) with PDB ID 4RES.¹⁴ The structure of ligands (**C1**, **C2**, and **C3**) were obtained from PubChem NCBI.²⁰ Furthermore, the NaKATPase in complex with ligand was minimised to remove the sterical clashes between ligands and receptor. Since NAK-ATPase is located in the membrane, then lipid bilayer system was assembled using CHARMM-GUI membrane generator²¹ with 0.07 M KCL in the water explicit solvent. Then, the complete structure was converted to amber PDB format using CharmmLipid2amber.py. Hydrogen atoms were added to the protein structure using Leap module of AMBER 14.²² The AM1-BCC partial atomic charges of ligands were calculated using antechamber module of AMBER 14. Minimisation was performed using Amber force field ff14SB,²³ General Amber Force Field, and Amber Lipid force field 14²⁴ using 250 steepest descents and 750 conjugate gradient algorithms. The minimised structure was converted to PDB format using ambpdb module of AMBER 14.

Protein-Ligand Docking and Psycho-chemical Property Calculations

Docking of all ligands was performed with AutoDock 4.2.²⁵ The structures of protein and ligand were extracted from the membrane and solvent molecules. Furthermore, they were converted into the pdbqt format by adding the atom type and charges using AutoDock Tools 1.5.6.²⁶ The K⁺ parameter was added to the AutoDock Parameter library, including its ionic charge. The size of the grid box of the receptor was 56 x 56 x 56

points with a grid spacing of 0.375 Å, centered in the binding site of bufalin. The Lamarckian Genetic algorithm was used with the following parameters: 100 number of runs, five million of energy evaluations, and 250 of populations. The energy breakdown analysis was performed using summary_result4.py program from AutoDockTools Utilities. The total binding energy was break down into hydrogen bond and electrostatic energies. In addition, the ligand efficiency (total binding energy divided by a number of atoms) of each ligand was calculated by this program. Psycho-chemical property such as PSA and A Log P have been computed by Biovia Draw 2016.²⁷

RESULT

The binding mode of bufalin in the NaK-ATPase was investigated from its crystal structure (PDB ID 4RES). It is shown that the 3-OH and 14-OH groups of bufalin formed hydrogen bonds with Glu117 and Thr797, respectively. The steroidal core of bufalin formed CH- π interaction with Phe783, while the rest of interactions between bufalin and NaK-ATPase was hydrophobic. Furthermore, all the bufadienolides (**C1**, **C2**, and **C3**) were superimposed to the crystal structure of bufalin inside the cavity of NaK-ATPase. It is shown that an oxygen atom of 1,3,5-orthoacetate moieties of **C1** and **C2** were in short distance with the carboxylate group of Glu117 (2.72 Å). Hence, a repulsive force was expected. Moreover, the 1-OH and 5-OH groups of **C3** were within the hydrogen bond distance with Glu117 (3.03 Å and 2.48 Å, respectively), while the aldehyde group (10-CHO) of **C1** and **C3** were predicted to form a hydrogen bond with Gln111 and Asn122. It is noted that the 14-OH group of all bufadienolides potentially formed a hydrogen bond with Thr797 Figure 2.

Protein-Ligand Docking and Psycho-chemical Property Calculations

A repulsive force between the Glu117 and the 1,3,5-orthoacetate group of **C1** and **C2** was predicted due to the sterical hindrance from a loop which Glu117 is located. For this reason, this loop was objected to a short minimisation scheme to avoid the unfavorable interaction with **C1** and **C2**. After minimisation, the interaction between 1,3,5-orthoacetate group and the loop was refined Figure 3. In general, the minimised structure of receptor has better interaction with all bufadienolides than the original crystal structure. Therefore, this structure was further used to dock all bufadienolides (**C1**, **C2**, and **C3**). As a result, there was a good correlation between the docking binding energy and the experimental data¹⁵ ($R^2 = 0.99$). Table 1 reveals that the **C1** has the lowest binding energy, followed by **C2** and **C3**. Regarding the polar surface area (PSA) and log P, all compounds were predicted to have good absorptivity properties Table 1. However, it is noted that the PSA and log P values of **C1** were better than the **C2** and **C3**. This ranking might also have explained the best activity of **C1** as compared to the others, due to a fact that NaKATPase is located in the lipid bilayer membrane.

Further interaction analysis using BIOVIA Discovery Studio Visualizer showed that the **C1** has four hydrogen bonds, four hydrophobic and seven CH- π interactions, while the **C2** has four hydrogen bonds, three hydrophobic and seven CH- π interactions. Lastly, the **C3** has the lowest number of interactions with only three hydrogen bonds, three hydrophobic and five CH- π interactions with the receptor. The 10-CHO group of **C1** and **C3** formed hydrogen bonds with Asn122 and Gln111, respectively, while the 10-CH₂OH group of **C2** formed a hydrogen bond with Gln111. An oxygen atom at the position 1 of the 1,3,5-orthoacetate moiety in **C1** and **C2** and the 1-OH group of **C3** formed hydrogen bonds with Gln111. The 11-OH group of **C1** and **C2** formed hydrogen bonds with Asn122 and the backbone of Ile315, respectively. The 14-OH group of all bufadienolides formed hydrogen bonds with Thr797.

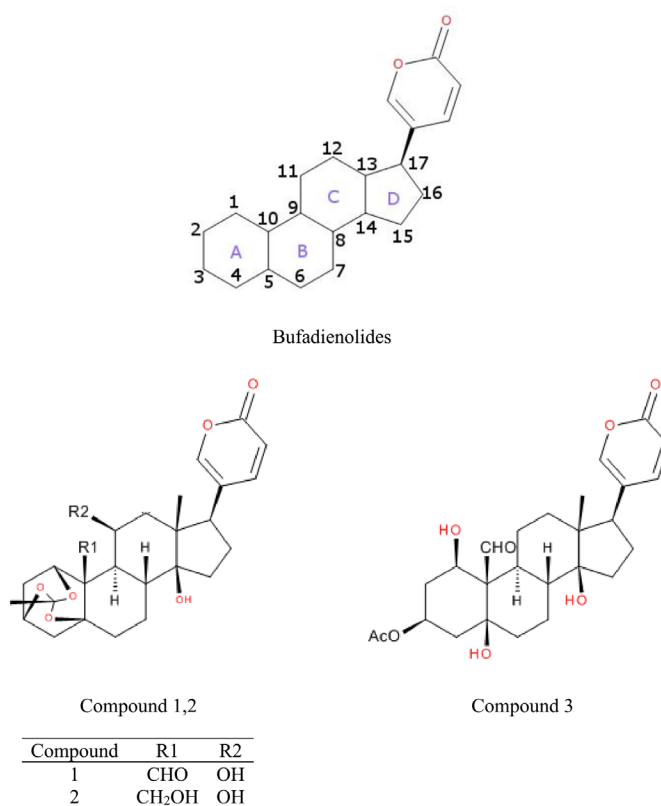


Figure 1: Structures of bufadienolides from *Kalanchoe pinnata*.

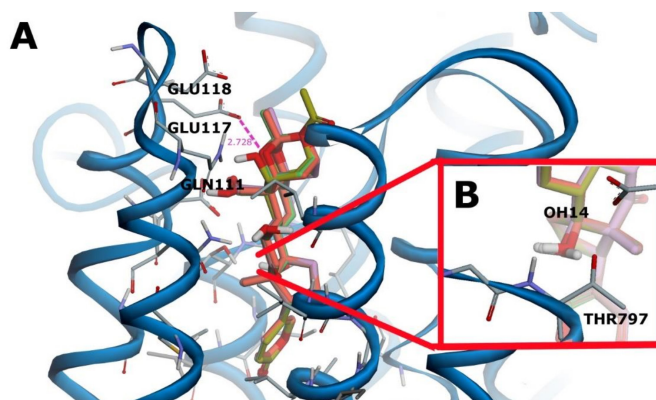


Figure 2: (A) Superimposed ligands in the binding cavity of NaK. An unfavorable interaction between the orthoacetate moiety and Glu117 was observed. (B) 14-OH of all bufadienolides potentially form hydrogen bond with Thr797.

Moreover, the energy breakdown analysis of docking result showed that **C1** has the highest ligand efficiency with strongest hydrogen bond and electrostatic interaction energies, followed by **C2** and **C3**. Table 2.

DISCUSSION

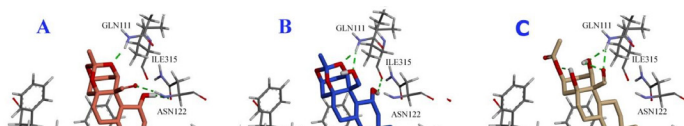
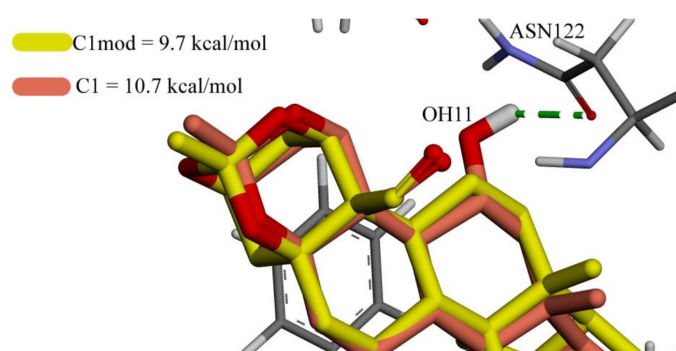
The previous study reported that a phorbol derivative induced the growth of tumor cell by over-stimulating the NaK-ATPase.⁵ Since a phorbol derivative was also used in Supratman's experiment,¹⁵ then it is suggested that the reduction of tumor cell growth by the bufadienolides was due to specific inhibition towards NaK-ATPase. The docking result

Table 1: Binding Energy and Experimental IC₅₀ of Bufadienolides from Kalanchoe Plants.

Compound	IC ₅₀ (μM)	Molecular Docking		Lipophilicity	
		RMSD (Å)	Binding Energy (kcal.mol ⁻¹)	PSA (Å ²)	Log P
C1	0.3	0.7	-10.7	111.5	0.9
C2	1.6	0.9	-9.5	114.7	0.7
C3	3.0	0.9	-8.9	130.4	0.4

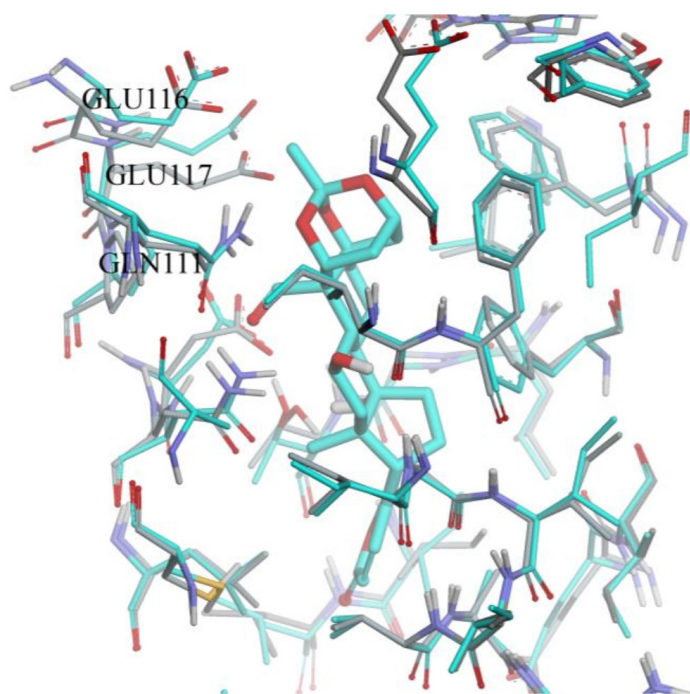
Table 2: Energy breakdown analysis and ligand efficiency of computed ligands.

Compound	Hydrogen bond energy (kcal/mol)	Electrostatic interaction energy (kcal/mol)	Ligand efficiency (Binding energy/Σatoms)
C1	-1.9	-0.3	-0.32
C2	-1.7	-0.1	-0.29
C3	-1.1	-0.1	-0.26

**Figure 3:** Crystal structure (grey) and minimisation structure (cyan) binding cavity of NaK. The residues seem to move further outside the binding cavity after minimisation process.**Figure 4:** Characteristic of hydrogen bond of CHO and CH₂OH at the position 10, (A) 10-CHO of C1 form hydrogen bond with ASN122, (B) 10-CH₂OH of C2 form hydrogen bond with GLN111, (C) 10-CHO of C3 form an intramolecular hydrogen bond with 1-OH and hydrogen bond with GLN111.

showed good correlation with the experimental value. The 1,3,5-orthoacetate moiety, 10-CHO, 11-OH, and 14-OH were suggested as the important substituents for antitumor activity of bufadienolide.

In this study, a crystal structure of NaK-ATPase which originated to genus *Sus scrofa* (PDB ID 4RES), with 98% identity with human NaK-ATPase, was used as a protein target. The molecular mechanics minimisation, besides removing the sterical hindrance from Glu117 to a 1,3,5-orthoacetate moiety, also relaxing the residues in the binding cavity and forming better interaction with bufadienolides.

**Figure 5:** The effect of 11-OH to the docking binding energy of C1 (red color) and C1mod (yellow color, the 11-OH is removed). Hydrogen bond is depicted in green-dashed line. C1mod loses a hydrogen bond with ASN122.

The 1,3,5-orthoacetate moiety is a distinctive feature of C1 and C2 as compared to the structure of C3 and the other common bufadienolides. In C3, substituents at position 1 and 5 are the OH-groups, while at position 3 is an acetate group. Mijatovic *et al.* suggested that the substituents in positions 1, 3, and 5 were not very critical to the inhibition activity.¹ However, when we introduced a 1,3,5-orthoacetate moiety to C3 and docked it to the NaK-ATPase, the binding affinity was improved.

Mijatovic¹ also suggested that the polar contact of the 10-CHO group with the Gln111 and Asn122 was not essential to the optimum interaction with NaK-ATPase. However, Larsen²⁸ showed that the 10-CHO

group formed a hydrogen bond with Gln111 and Asn122. From our results, we agreed that the presence of 10-CHO group and the 1,3,5-orthoacetate moiety lowered the binding energy by forming a hydrogen bond with Asn122. It is noted that the 10-CHO group in **C3** and 10-CH₂OH group in **C2** were not formed a hydrogen bond with Asn122, thus decreasing the inhibitory effect. The importance of asparagine at position 122 was tested by Canfield³⁰ and Lingrel.³¹ It is shown that the mutations of asparagine to glutamic acid at the position 122 (N122E) in the NaK-ATPase decreased the inhibition activity of CS (ouabain). Therefore, this agrees with our proposal of the importance of hydrogen bond with Asn122, despite the simplified approach used in docking method.

The role of 11-OH group to the binding energy was observed in this study. We purposely removed the 11-OH from **C1** compound Figure 5, which decreased the binding affinity by losing a hydrogen bond with Asn122. In contrast, Kamano *et al.* suggested that the 11-OH was not an important substituent for ligand binding.²⁹ Interestingly, Supratman *et al.* found that the 11-OH group decreased the cytotoxicity of bufadienolides.¹⁵ Thus, it is indicated that the formation of hydrogen bonds with Ile315 (in **C2**) and Asn122 (in **C1**) would increase the ligand's selectivity to NaK-ATPase. Therefore, a hydrogen bond with Asn122 might be one of the reasons for the better activity of **C1** as compared to the **C2**. Furthermore, the energy breakdown analysis showed that **C1** has the highest ligand efficiency among all, which means that the individual interaction in **C1** was stronger than **C2**. Despite the similar number of hydrogen bond formed by **C1** and **C2**, the 1,3,5-orthoacetate moiety and 10-CH₂OH in **C2** shared the hydrogen bonds with Gln111 Figure 4. This sharing interaction would result in a weaker hydrogen bond,³² and hence the total energy of hydrogen bond in **C2** was less than that of **C1**.

Regarding the physico-chemical properties of **C1-C3**, it is shown that their PSA and Log P values were correlated with their binding affinity, i.e., hydrophobicity increased the activity. As shown in Table 1, all of the Log P and PSA values were less than 5 and 140, respectively, indicating a good membrane permeability property.^{33,34} This result suggested that bufadienolides compounds, especially **C1**, would be able to enter the hydrophobic binding cavity of NaK-ATPase.

CONCLUSION

In summary, we have performed molecular docking to study the structure-activity relationship of bufadienolides derivatives from *K. pinnata* to the inhibition of NaK-ATPase. The crystal structure of NaK-ATPase from *Sus scrofa* bound to bufalin was used as protein target and as a control positive. The docking result showed a good agreement between the calculated binding energy and the experimental data. The presence of 1,3,5-orthoacetate moiety, 10-CHO, 11-OH, and 14-OH increased the activity of **C1** than the other bufadienolides. Although the role of 11-OH group was still debatable, this substituent appeared to decrease the toxicity of the bufadienolide and hence better selectivity. The result is also supported by the psycho-chemical properties value (PSA and LogP). This study suggests that **C1-C3** possess good selectivity to NaK-ATPase and low toxicity, thus interesting to be further studied for anticancer activity.

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REFERENCES

- Mijatovic T, Dufasne F, Kiss R. Cardiotonic steroids-mediated targeting of the Na⁺/K⁺-ATPase to combat chemo resistant cancers. *Current Medicinal Chemistry*. 2012;19(5):627-46.
- Johnson PP, Walker R, Jones SW, Stephens K, Meurer J, Zajchowski Da, *et al.* Multiplex gene expression analysis for high-throughput drug discovery: Screening and analysis of compounds affecting genes overexpressed in cancer cells. *Molecular Cancer Therapeutics*. 2002;1(14):1293-304.
- Mijatovic T, Van-Quaquebeke E, Delest B, Debeir O, Darro F, Kiss R. Cardiotonic steroids on the road to anti-cancer therapy. *Biochemical Biophysical Acta*. 2007;1776(1):32-57.
- Prassas I, Diamandis Ep. Novel therapeutic applications of cardiac glycosides. *Nature Reviews Drug Discovery*. 2008;7(11):926-35.
- Mijatovic T, Dufasne F, Kiss R. Na⁺/K⁺-ATPase and cancer. *Pharmaceutical Patent Analyst*. 2012;1(1):91-106.
- Mijatovic T, Kiss R. Cardiotonic steroids-mediated Na⁺/K⁺-ATPase targeting could circumvent various chemo resistance pathways. *Planta Med*. 2013;79(03/04):189-98.
- Efferth T, Davey M, Olbrich A, Rücker G, Gebhart E, Davey R. Activity of drugs from traditional Chinese medicine toward sensitive and Mdr1- Or Mrp1-Over-expressing multidrug-resistant human ccrf-cem leukemia cells. *Blood Cells, Molecules and Diseases*. 2002;28(2):160-8.
- Wei X, Si N, Zhang Y, Zhao H, Yang J, Wang H, *et al.* Evaluation of bufadienolides as the main antitumor components in cinobufacin injection for liver and gastric cancer therapy. *PLoS one*. 2017;12(1):e0169141.
- Newman RA, Yang P, Pawlus AD, Block KI. Cardiac glycosides as novel cancer therapeutic agents. *Molecular Intervention*. 2008;8(1):36-49.
- Gao H, Popescu R, Kopp B, Wang Z. Bufadienolides and their antitumor activity. *Nat Prod Rep*. 2011;28(5):953-69.
- Xie Z, Cai T. Na⁺-K⁺-ATPase-mediated signal transduction: From protein interaction to cellular function. *Molecular Interventions*. 2003;3(3):157-68.
- Juel C, Nordsborg Nb, Bangsbo J. Purinergic effects on Na,K-ATPase activity differ in rat and human skeletal muscle. *Plos One* 2014;9(3):E91175.
- Babula P, Masarik M, Adam V, Provaznik I, Kizek R. From Na⁺/K⁺-ATPase and cardiac glycosides to cytotoxicity and cancer treatment. *Anticancer Agents Med Chemistry*. 2013;13(7):1069-87.
- Laursen M, Gregersen JI, Yatime L, Nissen P, Fedosova Nu. Structures and characterization of digoxin- and bufalin-bound na⁺, k⁺-atpase compared with the ouabain-bound complex. *Proc Natl Academy Sci*. 2015;112(6):1755-60.
- Supratman U, Fujita T, Akiyama K, Hayashi H, Murakami A, Ohigashi H, *et al.* Anti-Tumor promoting activity of bufadienolides from *Kalanchoe Pinnata* and *k. daigremontiana x tubiflora*. *Bioscience Biotechnology and Biochemistry* 2001;65(4):947-9.
- Gago F. Molecular simulations of drug-receptor complexes in anticancer research. *Future Med Chemistry*. 2012;4(15):1961-70.
- Marco E, Laine W, Tardy C, Lansiaux A, Iwao M, Ishibashi F, *et al.* Molecular determinants of topoisomerase I am poisoning by lamellarins: Comparison with camptothecin and structure-activity relationships. *Journal of Medicinal Chemistry*. 2005;48(11):3796-807.
- Baskaran C, Ramachandran M. Computational molecular docking studies on anticancer drugs. *Asian Pacific Journal of Tropical Disease*. 2012;2:S734-8.
- Kingsley B, Kumari S, Subramoniam A, Pemiah B. In silico Docking studies on ATP-sensitive K⁺-channel, insulin receptor and phosphorylase kinase activity by isolated active principles of *stereospermum tetragonum* DC. *J Young Pharm*. 2017;9(1):124-6.
- Kim S, Thiessen PA, Bolton EE, Chen J, Fu G, Bryant SH, *et al.* PubChem Substance and Compound databases. *Nucleic Acids Res*. 2015;44(D1):D1202-13.
- Wu EL, Cheng X, Jo S, Rui H, Song KC, Dávila-Contreras EM, *et al.* CHARMM-GUI Membrane Builder Toward Realistic Biological Membrane Simulations. *J Computational Chemistry*. 2014;35(27):1997-2004.
- Case DA, Babin V, Berryman JT, Betz RM, Cai Q, Kollman PA, *et al.* AMBER 14. University of California, San Francisco. 2014.
- Maier JA, Martinez C, Kasavajhala K, Wickstrom L, Hauser KE, Simmerling C. ff14SB: improving the accuracy of protein side chain and backbone parameters from ff99SB. *J Chemical Theory Computation*. 2015;11(8):3696-713.
- Dickson CJ, Madej BD, Skjevik AA, Betz RM, Teigen K, Gould IR, *et al.* Lipid14: The Amber lipid force field. *J Chemical Theory Computation*. 2014;10(2):865-79.
- Morris GM, Huey R, Lindstrom W, Sanner MF, Belew RK, Goodsell DS, Olson AJ. Autodock4 and AutoDockTools4: automated docking with selective receptor flexibility. *J Computational Chemistry*. 2009;30(16):2785-91.
- Michel FS. Python: A Programming Language for Software Integration and Development. *J Molecular Graphics Mod*. 1999;17(1):57-61.
- Biovia Draw 2016. Accelrys Technologies, Inc. <http://accelrys.com/products/collaborative-science/biovia-draw>.
- Laursen M, Yatime L, Nissen P, Fedosova NU. Crystal structure of the high-affinity Na⁺/K⁺-ATPase-ouabain complex with Mg²⁺ bound in the cation binding site. *Proc Natl Academy Sci USA*. 2013;110(27):10958-63.
- Canfield V, Emanuel JT, Spickofsky N, Levenson R, Margolskeel RF. Ouabain-Resistant mutants of the Rat Na, K-ATPase α2 isoform identified by using an episomal expression vector. *Molecular and Cellular Biology*. 1990;10(4):1367-72.

30. Lingrel JB, Arguello JM, Van-Huysse J, Kuntzweiler TA. Cation and cardiac glycoside binding sites of the Na, K-ATPase. *Ann NY Academy Sci.* 1997;834(1):194-206.
31. Kamano Y, Kotake A, Hashima H, Inoue M, Morita H, Takeya K, *et al.* Structure-Cytotoxic Activity Relationship for the toad poison bufadienolides. *Bioorganic and Medicinal Chemistry.* 1998;6(7):1103-15.
32. Hydrogen Bond in Water. http://www1.lsbu.ac.uk/water/water_hydrogen_bonding.html
33. Lipinski CA, Lombardo F, Dominy BW, Feeney PJ. Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings. *Adv Drug Delivery Rev.* 1997;23(1-3):3-25. (2001;46(1-3):3-26)
34. Pajouhesh H, Lenz GR. Medicinal chemical properties of successful central nervous system drugs. *NeuroRx.* 2005;2(4):541-53.

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